

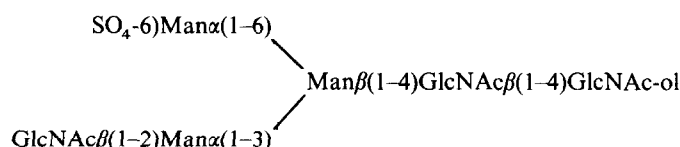
Primary structure of the acidic carbohydrate chain of hemocyanin from *Panulirus interruptus*

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The *N*-glycosidic carbohydrate chains of hemocyanin from the spiny lobster *Panulirus interruptus* were liberated by hydrazinolysis of a pronase digest and subsequently reduced. Separation of the mixture of oligosaccharide-alditols by high-voltage paper electrophoresis resulted in a neutral (90%) and an acidic (10%) fraction. 500-MHz ^1H -NMR spectroscopy of the acidic fraction revealed a single component with the following novel structure:



Hemocyanin; *N*-linked carbohydrate; Sulfate; (*Panulirus interruptus*)

1. INTRODUCTION

Hemocyanins are high- M_r copper-containing oxygen-transport proteins, which are found in the hemolymph of some arthropods and molluscs. It has been shown that interesting species-specific differences in carbohydrate chains occur among the hemocyanins in both phyla [1-5]. The hemo-

cyanin from the spiny lobster *Panulirus interruptus* is an *N*-glycoprotein, having a carbohydrate content of 1.5% with Man and GlcNAc as the only constituting monosaccharides [5]. Recently, the primary structures of the neutral carbohydrate chains were established to be $\text{Man}\alpha(1\text{-}6)[\text{GlcNAc}\beta(1\text{-}2)\text{Man}\alpha(1\text{-}3)]\text{Man}\beta(1\text{-}4)\text{GlcNAc}\beta(1\text{-}4)\text{GlcNAc}$ and $\text{Man}\alpha(1\text{-}6)[\text{Man}\alpha(1\text{-}3)]\text{Man}\alpha(1\text{-}6)[\text{Man}\alpha(1\text{-}3)]\text{Man}\beta(1\text{-}4)\text{GlcNAc}\beta(1\text{-}4)\text{GlcNAc}$ [5]. Here, the structural characterization of the acidic carbohydrate material, representing 10% of the carbohydrate part, is described.

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Abbreviations: Man, D-mannopyranose; GlcNAc, 2-acetamido-2-deoxy-D-glucopyranose; GlcNAc-ol, 2-acetamido-2-deoxy-D-glucitol

2. MATERIALS AND METHODS

The preparation and isolation of the acidic carbohydrate fraction from a hydrazinolysate of the

pronase digest of *P. interruptus* hemocyanin have been described in [5].

Methyl α -D-mannopyranoside 6-*O*-sulfate was synthesized according to [6]. Briefly, methyl α -D-mannopyranoside (194 mg, 1 mmol) was dissolved in 5 ml dry pyridine. After cooling to 5°C, 73 μ l chlorosulfonic acid (1.1 mmol) in 300 μ l dry chloroform was added. The mixture was stirred for 30 min at 5°C, and then for 2 h at 25°C. After the addition of 2 ml water, the solvent was evaporated to dryness. The crude material was purified on a silica column (2 $\times$ 7 cm), using a mixture of dichloromethane:methanol (5:3, v/v) as eluting system. The final product was converted into the corresponding sodium salt by neutralization with NaOH.

For $^1\text{H-NMR}$ analysis, samples were repeatedly exchanged in $^2\text{H}_2\text{O}$ (99.96 atom% ^2H , Aldrich) with intermediate lyophilization. $^1\text{H-NMR}$ spectra were recorded on a Bruker AM-500 spectrometer (SON hf-NMR facility, Department of Biophysical Chemistry, University of Nijmegen, The Netherlands) operating at 500 MHz in the Fourier-transform mode at a probe temperature of 27°C. One-dimensional spin lock difference spectroscopy was performed according to [7], in combination with the MLEV-17 composite pulse decoupling cycle [8], using a spin lock time of 160 ms. Resolution enhancement of the spectra was achieved by Lorentzian-to-Gaussian transformation [9]. Chemical shifts (δ) are given relative to sodium 4,4-dimethyl-4-silapentane-1-sulfonate, but were actually measured indirectly to acetone in $^2\text{H}_2\text{O}$ (δ 2.225) [10].

3. RESULTS AND DISCUSSION

Hydrazinolysis of a pronase digest of hemocyanin from the spiny lobster *P. interruptus* resulted in complete liberation of the *N*-linked carbohydrate chains [5]. After re-*N*-acetylation and reduction, the mixture of oligosaccharide-alditols was fractionated by high-voltage paper electrophoresis, yielding 10% acidic material, which chromatographed as one peak, denoted **A**. In view of the sugar analysis of the native glycoprotein, showing only Man and GlcNAc as monosaccharides, no sugar constituent can be held responsible for the acidic nature of fraction **A**.

To elucidate the primary structure of the oligo-

saccharide-alditol present in fraction **A**, a 500-MHz $^1\text{H-NMR}$ spectrum was recorded (fig.1A). Relevant $^1\text{H-NMR}$ data of **A**, together with those of $\text{Man}\alpha(1\text{--}6)[\text{GlcNAc}\beta(1\text{--}2)\text{Man}\alpha(1\text{--}3)]\text{Man}\beta(1\text{--}4)\text{GlcNAc}\beta(1\text{--}4)\text{GlcNAc-ol}$, being the previously reported structure in the neutral fraction **c** [5], are presented in table 1. The equal intensity of the anomeric proton signals in the $^1\text{H-NMR}$ spectrum of **A** points to the presence of a single compound. When comparing the $^1\text{H-NMR}$ data of **A** and **c** (table 1), it is evident that all structural-reporter groups which are present in the spectrum of **c** also occur in that of **A**, having essentially the same chemical shifts. This means that the compound in fraction **A** and **c** must have the structural element $\text{GlcNAc}\beta(1\text{--}2)\text{Man}\alpha(1\text{--}3)[\text{Man}\alpha(1\text{--}6)]\text{Man}\beta(1\text{--}4)\text{GlcNAc}\beta(1\text{--}4)\text{GlcNAc-ol}$ in common. However, the spectrum of **A** shows two additional downfield signals, resonating at δ 4.354 and δ 4.2. The doublet of doublets at δ 4.354 has a large coupling constant of 11.6 Hz, which can only correspond to a geminal coupling of H-6 and H-6' of Man, or GlcNAc. To identify this monosaccharide residue, one-dimensional spin lock difference spectroscopy was performed. For this purpose a selective 180° pulse was given to the signal at δ 4.354 and the difference spectrum revealed the complete set of proton signals from one sugar unit (fig.1B). This residue could be identified as Man-4' from the H-2 signal at δ 3.979, and from the (less distinct) H-1 signal at δ 4.911. The unknown signal at δ 4.2 belongs to H-6' of Man-4'. The appearance of the Man-4' H-6 and H-6' at downfield positions as found for **A**, in comparison to **c**, must be attributed to the presence of an acidic substituent at C-6. In view of the literature data on acidic carbohydrate chains, it is reasonable to propose that either a phosphate or a sulfate group is involved. It has been demonstrated that 6-*O*-phosphorylation of a terminal α -D-Man residue causes a downfield shift of H-6 and H-6' to δ 4.061 in $\text{Man}\alpha(1\text{--}6)\text{Man-R}$ [11] and a downfield shift of H-6 to δ 4.096 in $\text{Man}\alpha(1\text{--}2)\text{Man}$, whereby a $^1\text{H-}^{31}\text{P}$ coupling (6.5 Hz) is evident [12]. However, in the $^1\text{H-NMR}$ spectrum of **A**, the H-6 (and H-6') are shifted to even more downfield positions than observed for H-6 (and H-6') in terminal 6-*O*-phosphorylated α -D-mannose residues [11,12]. Furthermore, no $^1\text{H-}^{31}\text{P}$ coupling is present on the Man-4' H-6 and H-6' signals. Therefore, a sulfate

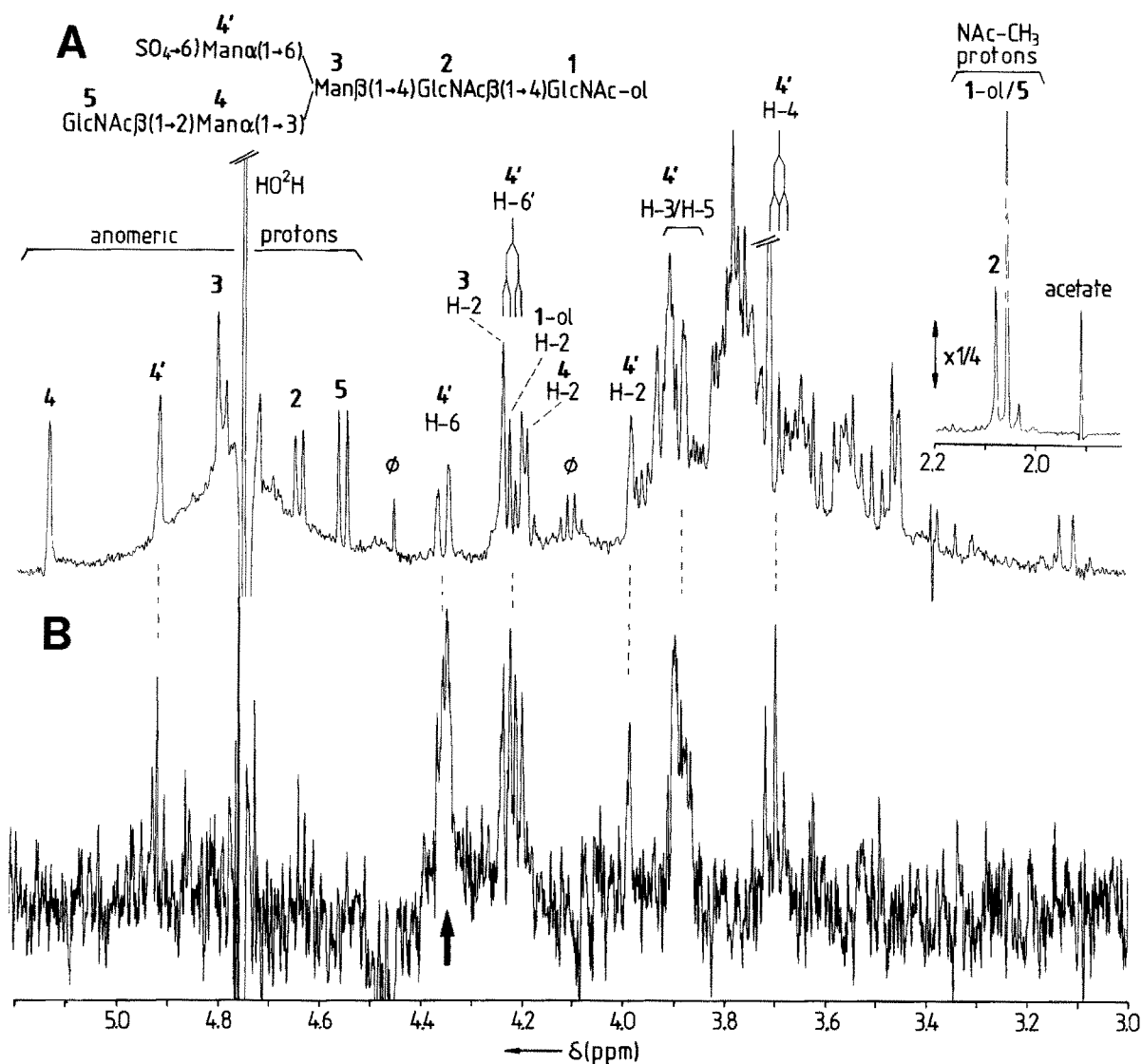


Fig.1. (A) Structural-reporter group regions of the resolution-enhanced 500-MHz ^1H -NMR spectrum ($^2\text{H}_2\text{O}$ at 27°C) of oligosaccharide-[1- ^2H]alditol fraction **A** from *P. interruptus* hemocyanin. The numbers in the spectrum refer to the corresponding residues in the structure. The relative intensity scale of the *N*-acetyl region differs from that of the other part of the spectrum as indicated. ϕ denotes impurity. (B) Spin lock difference spectrum, with a selective 180° pulse (indicated by the arrow) on the H-6 signal of Man-4'.

group should be present. To confirm this suggestion, the ^1H -NMR spectra of the methyl α -D-mannopyranoside and methyl α -D-mannopyranoside 6-*O*-sulfate were recorded. As is evident from table 2, sulfation at C-6 causes a downfield shift of H-6 and H-6' to δ 4.344 and 4.229, respectively. These chemical shift values are of the same order

as those observed for Man-4' H-6 (δ 4.354) and H-6' (δ 4.218) in **A**. It should be noted that the H-6 and H-6' signals of Man-4' are essentially the same as those described for terminal GlcNAc, *O*-sulfated at C-6 (H-6, δ 4.337; H-6', δ 4.219) in GlcNAc β (1-3)Gal β [13]. Summarizing the various data, the structure of **A** is proposed to be:

Table 2

¹H-NMR chemical shifts for the methyl α -D-glycopyranosides of mannose and mannose 6-O-sulfate

Protons	Chemical shift ^a (ppm)	
	Man	Man-6-S
H-1	4.761	4.763
H-2	3.929	3.938
H-3	3.751	3.626
H-4	3.640	3.700
H-5	3.604	3.824
H-6	3.898	4.344
H-6'	3.755	4.229
OCH ₃	3.407	3.418

limited. Some details have been reported for 4-*O*-sulfated Man in the carbohydrate chains of hen egg albumin [14] and 3/4-*O*-sulfated GalNAc in the carbohydrate chains of lutropin [15]. Man-6-sulfate has been discovered recently as a constituent of the *N*-linked carbohydrate chains of the lysosomal enzymes from *Dictyostelium discoideum* [16].

^b Chemical shifts are given for neutral solutions at 27°C, in ppm downfield from sodium 4,4-dimethyl-4-silapentane-1-sulfonate in ²H₂O acquired at 500 MHz (but were actually measured relative to internal acetone: δ 2.225)

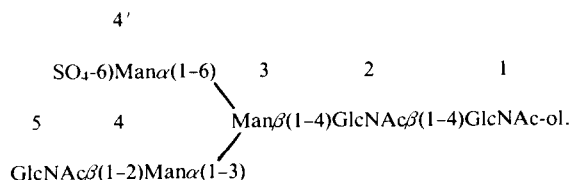
^d n.d., not determined

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Primary structural studies on sulfated *N*-linked carbohydrate chains have thus far been very

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